

FORM PTO-1390 (Modified)
(REV 11-2000)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

TRANSMITTAL LETTER TO THE UNITED STATES

HER0050

DESIGNATED/ELECTED OFFICE (DO/EO/US)

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR

CONCERNING A FILING UNDER 35 U.S.C. 371

10/031477

INTERNATIONAL APPLICATION NO.

PCT/FR00/02072

INTERNATIONAL FILING DATE

JULY 19, 2000

PRIORITY DATE CLAIMED

JULY 20, 1999

TITLE OF INVENTION

USE OF PROPIONIC BACTERIA FOR PRODUCING PROPIONIC ACID AND/OR PROPIONATES IN THE COLON

APPLICANT(S) FOR DO/EO/US

EDMOND DANIEL ROUSSEL, ET AL.

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☐ This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (24) indicated below.
4. ☒ The US has been elected by the expiration of 19 months from the priority date (Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371 (c) (2))
 - a. ☐ is attached hereto (required only if not communicated by the International Bureau).
 - b. ☒ has been communicated by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☒ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
 - a. ☒ is attached hereto.
 - b. ☐ has been previously submitted under 35 U.S.C. 154(d)(4).
7. ☐ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))
 - a. ☐ are attached hereto (required only if not communicated by the International Bureau).
 - b. ☐ have been communicated by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☐ have not been made and will not be made.
8. ☐ An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☐ An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).
10. ☐ An English language translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).
11. ☒ A copy of the International Preliminary Examination Report (PCT/IPEA/409).
12. ☒ A copy of the International Search Report (PCT/ISA/210).

Items 13 to 20 below concern document(s) or information included:

13. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
14. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. ☒ A **FIRST** preliminary amendment.
16. ☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
17. ☐ A substitute specification.
18. ☐ A change of power of attorney and/or address letter.
19. ☐ A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821 - 1.825.
20. ☐ A second copy of the published international application under 35 U.S.C. 154(d)(4)
21. ☐ A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).
22. ☒ Certificate of Mailing by Express Mail
23. ☐ Other items or information:

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR 1.01) <div style="font-size: 1.5em; font-weight: bold;">10/031477</div>		INTERNATIONAL APPLICATION NO <div style="font-weight: bold;">PCT/FR00/02072</div>		ATTORNEY'S DOCKET NUMBER <div style="font-weight: bold;">HER0050</div>	
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24. The following fees are submitted: <div style="margin-top: 10px;"> BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)) : <div style="margin-left: 20px;"> <input type="checkbox"/> Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO \$1040.00 </div> <div style="margin-left: 20px;"> <input checked="" type="checkbox"/> International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$890.00 </div> <div style="margin-left: 20px;"> <input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$740.00 </div> <div style="margin-left: 20px;"> <input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$710.00 </div> <div style="margin-left: 20px;"> <input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) \$100.00 </div> </div>				CALCULATIONS PTO USE ONLY	
ENTER APPROPRIATE BASIC FEE AMOUNT =				\$890.00	
Surcharge of \$130.00 for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492 (e)).				\$0.00	
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE		
Total claims	9 - 20 =	0	x \$18.00	\$0.00	
Independent claims	1 - 3 =	0	x \$84.00	\$0.00	
Multiple Dependent Claims (check if applicable). <input type="checkbox"/>				\$0.00	
TOTAL OF ABOVE CALCULATIONS =				\$890.00	
<input checked="" type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2.				\$445.00	
SUBTOTAL =				\$445.00	
Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492 (f)).				\$0.00	
TOTAL NATIONAL FEE =				\$445.00	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable). <input type="checkbox"/>				\$0.00	
TOTAL FEES ENCLOSED =				\$445.00	
				Amount to be: refunded \$	
				charged \$	

a. ☒ A check in the amount of **\$445.00** to cover the above fees is enclosed.

b. ☐ Please charge my Deposit Account No. _____ in the amount of _____ to cover the above fees. A duplicate copy of this sheet is enclosed.

c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. **02-0385**. A duplicate copy of this sheet is enclosed.

d. ☐ Fees are to be charged to a credit card. **WARNING:** Information on this form may become public. **Credit card information should not be included on this form.** Provide credit card information and authorization on PTO-2038.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

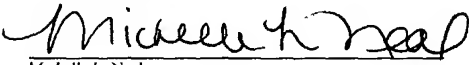
Anthony Niewyk, Reg. No. 24,871
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SIGNATURE
 Anthony Niewyk
 NAME
 24,871
 REGISTRATION NUMBER
 January 18, 2002
 DATE

10/031477
JC13 Rec'd PCT/PTO 18 JAN 2002

PATENT APPLICATION

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

<i>Group</i>		}	Certificate Under 37 C.F.R. 1.10
<i>Art Unit:</i>	Unknown	}	"EXPRESS MAIL" MAILING LABEL NUMBER
		}	EL731285032US
<i>Attorney</i>		}	DATE OF DEPOSIT <u>January 18, 2002</u>
<i>Docket No.:</i>	HER0050	}	I HEREBY CERTIFY THAT THIS PAPER OR FEE IS BEING
		}	DEPOSITED WITH THE UNITED STATES POSTAL SER-
<i>Applicant:</i>	Edmond D. Roussel et al.	}	VICE "EXPRESS MAIL POST OFFICE TO ADDRESSEE"
		}	SERVICE UNDER 37 C.F.R. 1.10 ON THE DATE
<i>Invention:</i>	USE OF PROPIONIC BACTERIA FOR	}	INDICATED ABOVE AND IS ADDRESSED TO THE ASSIS-
	PRODUCING PROPIONIC ACID	}	TANT COMMISSIONER FOR PATENTS WASHINGTON, DC
	AND/OR PROPIONATES IN THE	}	20231
	COLON	}	on <u>January 18, 2002</u>
		}	
		}	Michelle L. Neal
<i>Serial No:</i>	Unknown	}	
<i>Filed:</i>	Herewith	}	
<i>Internat'l Serial</i>	PCT/FR00/02072	}	
<i>Internat'l Filing</i>		}	
<i>Date:</i>	July 19, 2000	}	
<i>Earliest Priority</i>		}	
<i>Date:</i>	July 20, 1999	}	
<i>Examiner:</i>	Unknown	}	

PRELIMINARY AMENDMENT

Box PCT
Attn: DO/EO/US
Assistant Commissioner for Patents
Washington, DC 20231

Sir:

Prior to the examination of the above-identified application, please amend the application
as follows:

IN THE CLAIMS

Please amend claims 4-9 as follows:

4. (Amended) Use as claimed in Claim 1, characterized in that the propionic bacteria belong to strains having properties of adhesion on the colonocytes.

5. (Amended) Use as claimed in Claim 1, characterized in that the composition consists of a dry or hydrated preparation presented in the form of individual fractions of approximately 100 mg to 1 g, preferably from 200 to 500 mg, preferably containing at least 10^8 cells.

6. (Amended) Use as claimed in Claim 5, characterized in that the composition is presented in the form of gastroresistant capsules.

7. (Amended) Use as claimed in Claim 1, characterized in that the composition consists of a formulated preparation, the propionic bacteria being added to or associated with a fermentable substrate, notably dietary fibers.

8. (Amended) Use as claimed in Claim 1, characterized in that the composition consists of a formulated preparation, the propionic bacteria being added to or incorporated into liquid, paste or solid foods.

9. (Amended) Use as claimed in Claim 1, characterized in that the composition contains lactic bacteria and/or bifid bacteria.

• • • R E M A R K S • • •

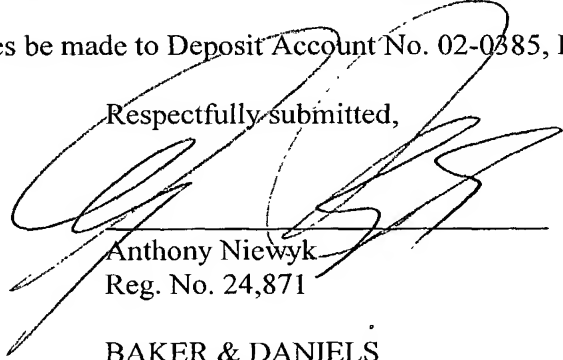
By the present Preliminary Amendment the claims have been revised delete the multiple dependency.

Care has been taken so as to avoid the addition of new matter in the claims.

Entry of the present Preliminary Amendment prior to the examination of the application is respectfully requested.

In the event applicants have overlooked the need for an extension of time, an additional extension of time, payment of fee, or additional payment of fee, applicants hereby petition therefor and authorize that any charges be made to Deposit Account No. 02-0385, Baker & Daniels.

Respectfully submitted,



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Claim 1 remains unchanged.

Claim 3 remains unchanged.

4. (Amended) Use as claimed in [any one of Claims 1 to 3] Claim 1, [characterised]
characterized in that the propionic bacteria belong to strains having properties of adhesion on the
colonocytes.

5. (Amended) Use as claimed in [any one of Claims 1 to 4] Claim 1, [characterised]
characterized in that the composition consists of a dry or hydrated preparation presented in the form
of individual fractions of approximately 100 mg to 1 g, preferably from 200 to 500 mg, preferably
containing at least 10^8 cells.

6. (Amended) Use as claimed in Claim 5, [characterised] characterized in that the composition is presented in the form of gastroresistant capsules.

Claim 7 has been amended as follows:

7. (Amended) Use as claimed in [any one of Claims 1 to 6] Claim 1, [characterised] characterized in that the composition consists of a formulated preparation, the propionic bacteria being added to or associated with a fermentable substrate, notably dietary [fibres] fibers.

Claim 8 has been amended as follows:

8. (Amended) Use as claimed in [any one of Claims 1 to 6] Claim 1, [characterised] characterized in that the composition consists of a formulated preparation, the propionic bacteria being added to or incorporated into liquid, paste or solid foods.

Claim 9 has been amended as follows:

9. (Amended) Use as claimed in [any one of Claims 1 to 7] Claim 1, [characterised] characterized in that the composition contains lactic bacteria and/or bifid bacteria.

1/pts

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The present invention relates to the use of propionic bacteria with a view to optimising the production of propionic acid and/or of propionates and, as the case may be, of acetic acid and/or of acetates at the colon.

For some years specialists in nutrition have been recommending to their patients a diet rich in fibres to which they attribute physiological and metabolic effects which may be beneficial to health.

It is known that dietary fibres resist the enzymatic digestion in the small intestine and are only degraded and assimilated at the colon, that is to say the terminal part of the intestine. Therefore the beneficial effect mentioned above can only be exerted if this degradation and this assimilation are as complete as possible at this preterminal location: the colon.

It has been possible to establish that these biological reactions result from the anaerobic fermentation of the dietary fibres under the action of micro-organisms in the colon. This fermentation culminates in the production of short-chain fatty acids, hydrogen, carbon dioxide and biomass.

These short-chain fatty acids are essentially acetic acid, propionic acid and butyric acid; in the healthy organism they can only be produced at the colon, since that is the only location in the human body where strict anaerobic conditions prevail which permit the fermentation at the base of their synthesis, with the exception of acetic acid of which a very small quantity may be produced in the hepatic region.

Different studies have proved the importance of these short-chain fatty acids which are beneficial to health.

According to the literature it would appear that the physiological roles of these three short-chain fatty acids would be different from one another: in effect, the acetic and propionic acids would be led directly towards the liver where the totality of the propionic acid would be

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metabolised whilst a part of the acetic acid would then be led to different tissues, whilst the butyric acid would be used more specifically within the wall of the colon.

Therefore the synthesis of the short-chain fatty acids implies the presence in the colon, on the one hand, of a fibre-based substrate which is easy to provide through food and, on the other part, of a balanced and adapted bacterial flora which is present in an optimal and constant manner.

This bacterial flora can originate from either the persistent endogenous flora of each individual or from the diet.

It is in fact well known that the contents of the human digestive tract, which is specific to each individual and corresponds approximately to 1 to 1.5 kg of food material in the course of digestive transformation, contains a significant population of micro-organisms consisting of a mixture of numerous species which may be evaluated at 10^{11} to 10^{12} cells per gram in the colon; this population constitutes a bacterial mass of a certain weight of which the good or bad balance can only be modified radically and above all durably with difficulty through the simple fact of current diet.

However, the food which one ingests daily is never sterile and is therefore more or less charged with bacteria (milk, fermented milk products, cheese, cider, wine, beer, meat, etc.). Nevertheless, the modifications to the colic flora as a result of the absorption of these bacteria can only be temporary.

It may also be noted that it has already been proposed to attempt to modify the microbial population of the intestinal tract by the administration and in particular the voluntary ingestion of bacterial cells reputed to be beneficial to health (known as probiotics), particularly lactic bacteria or bifid bacteria.

The introduction into the organism of a significant population of these bacteria either by means of a particular diet or by the direct ingestion of these microbial cells has been proposed

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more particularly with the aim of limiting the development of pathogenic and putrefying species; in fact it is known that the endogenous flora present in the colon is distributed into different bacterial groups of which certain are not harmful, and are indeed beneficial, whilst others, particularly clostridium and putrefying ones, lead to the production of toxic substances and have a negative influence on health.

The idea on which the invention is based consisted of introducing regularly into the organism, by the oral route, a significant quantity of a probiotic microbial flora capable of favouring the regular synthesis of short-chain fatty acids at the colon.

Amongst the microbial species which can be employed to this effect, lactic bacteria are not very well adapted since by their nature they produce first and foremost lactic acid and very secondarily acetic acid but not propionic acid nor butyric acid.

By contrast, bacteria of another type, the propionic bacteria, are capable of producing abundant quantities of propionic acid and acetic acid, the two short-chain fatty acids which are required to irrigate the tissue networks, for example in a percentage of 2/3 propionic acid to 1/3 acetic acid. These bacteria are present in cooked pressed cheeses; furthermore they have the advantage of being better equipped than the lactic bacteria to have an activity in the colon where the anaerobiosis is total and also of being more resistant to technological stresses than the lactic bacteria and the bifid bacteria.

It should be noted that it has already been proposed in the literature to cause the absorption of the propionic bacteria, in particular in order to stimulate the development of bifid bacteria in the intestine (document WO 97/19689) or to release nitrogen monoxide in the human or animal digestive tract (document WO 98/27991). However, until now no-one has ever had the idea of using these bacteria for the production of short-chain fatty acids at the colon.

Therefore the present invention relates to the use of propionic bacteria selected as a function of their nature which is not very autolytic and their ability to resist bile salts in order to obtain a current food composition or a dietetic or medicinal composition which is absorbable by a

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human or an animal, prepared so that the bacteria are protected at least partially against gastric acidity, containing at least 10^6 cells/gram of these bacteria, capable of stimulating and increasing significantly the synthesis of propionic acid and/or of propionate and, as the case may be, of acetic acid and/or of acetate at the colon by anaerobic bacterial fermentation.

In order that these bacterial should have the expected beneficial effect it is essential to choose strains which are not very autolytic which are capable of reaching the colon without damage, possibly developing there and producing sufficient quantities of propionic acid.

It is well known that the two principal stresses to which the ingested bacteria are subjected during their passage in the upper part of the digestive tract are associated on the one hand with the acidity of the stomach environment (pH 4 to 1) and on the other hand with the presence of bile salts in the small intestine (of the order of 15 mmol/l at most at the duodenum).

It has proved possible to establish that bacterial which have been exposed to the stomach acidity are weakened and consequently incapable of resisting the bile salts, even if they remain viable at the exit from the stomach.

Consequently, in accordance with the invention, it is essential to make the propionic bacteria undergo a treatment of a type which enables them not to undergo the gastric stress which as a general rule corresponds to an encapsulation which may be voluntary or involuntary for example in the case of a food of the cheese type.

This situation has been brought to light due to a test used to evaluate the influence of the acid pH and of the bile salts successively or individually on the viability of two strains of propionic milk bacteria belonging to the TL collection of the LRTL (Laboratoire de Recherches de Technologie Laitière – INRA of Rennes), namely the strains TL 162 and TL 24 belonging to the species *P. freudenreichii* subsp *shermanii*.

The results of this test are described below.



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The bacteria were cultivated at 30°C on a YEL medium for 2 days (start of stationary phase). The optical density at 650 nm was respectively 2.28 and 2.64 for TL 162 and TL 24.

- Acid stress

The cultures were diluted to 1/10th in S medium (tryptone-lactate) at pH 2.5 (final pH 3.0). After incubation at 37°C for 45 minutes the cultures were centrifuged and the bacteria taken up in the same volume of YEL. The counts were made before and at the end of the incubation, and the resumption of the growth was followed by measurements of DO for 5 days at 37°C.

- Bile stress

The initial cultures were centrifuged and the bacteria taken up in a volume 10 times greater of YEL containing 0.3% of bovine bile (~50% of bile salts). After an incubation at 37°C for 90 minutes, the cultures were centrifuged and the bacteria taken up in the same volume of YEL. Counts were made before and at the end of the incubation, and the resumption of the growth was followed by measurements of DO for 5 days at 37°C.

- Successive acid and bile stresses

The bacteria were subjected to an acid stress as described previously, but after centrifugation the cells were taken up in YEL containing 0.3% bile. After a second incubation at 37°C for 90 minutes, the cultures were centrifuged and the bacteria taken up in the same volume of YEL. Counts were made before and at the end of the incubation, and the resumption of the growth was followed by measurements of DO for 5 days at 37°C.

The results obtained are reported, on the one hand, in Table 1 below which mentions the influence of the acid and/or bile stress on the viability of the bacteria and, on the other hand, in Figure 1 which is a diagram representing the resumption of the growth after the different stresses.

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Table 1

		Viability (cfu/ml)		
		Before stress	After acid stress	After bile stress
Acid stress alone	TL 162	3.0×10^8	9.7×10^6	/
	TL 24	4.0×10^8	1.0×10^7	/
Bile stress alone	TL 162	3.0×10^8	/	4.4×10^8
	TL 24	4.0×10^8	/	4.7×10^8
Successive stresses	TL 162	3.0×10^8	9.7×10^6	2600
		4.0×10^8	1.0×10^7	< 10

It was also possible to establish that:

- the acidity involves a substantial mortality of the bacteria (96.8% for TL 162 and 97.5% for TL 124), which explains a sufficiently long delay for the resumption of the growth (Figure 1).
- the bile does not involve any mortality of the bacteria, hence a very rapid resumption of the growth.
- when the bacteria are subjected to the bile, after a previous acid stress, this leads to an almost total mortality of the bacteria. This result, totally unexpected, therefore indicates that bacteria which have undergone an acid stress and which nevertheless remain viable become totally sensitive to the bile, whilst without previous acid stress these same bacteria are totally resistant to the bile salts.

Taking this situation into account, pre-adaptation tests were carried out with the aim of increasing the resistance of the bacteria. It is known in fact that an acid pre-stress (pH 4.5 – 5) effectively protects the cells against an acid stress (pH 2).

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Therefore three adaptation tests were carried out on TL 162:

- *acid pre-stress*: preliminary incubation of the cells at 37°C for 30 minutes at pH 5

- *bile pre-stress*: incubation for 30 minutes in the presence of 0.08% of bile

- *acid and bile pre-stress*: incubation for 30 minutes at pH 5 and in the presence of 0.08% of bile.

The same protocol was applied as previously and the results obtained are reported in Table 2 below:

Table 2

	Viability (cfu/ml)	
	Before stress	After successive stresses
Without pre-adaptation	3×10^8	2600
Acid pre-adaptation	3×10^8	100
Bile pre-adaptation	3×10^8	1400
Acid and bile pre-adaptation	3×10^8	< 100

Thus it was possible to establish that an acid pre-adaptation weakens the cells even more, whilst a bile pre-adaptation has no effect.

Therefore these results have made it possible to demonstrate the necessity of treating the stresses successively and not separately as described in the majority of studies which have been carried out in this field.

However, it may be supposed that *in vivo* the conditions are less drastic for the bacteria (buffer effect of the food in the stomach, lesser bactericidal effect of the bile salts in micellar form with the phospholipids).

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Taking account of the foregoing, in order to increase the quantity of viable bacteria an improvement in their resistance to the acid pH is not effective since the bacteria remain sensitive to the effect of the bile salts.

On the other hand, by protecting the bacteria from the acid stress, in particular by ingesting them prepackaged in gastroresistant capsules, it is possible to find in the faeces bacteria which are naturally resistant to the bile at a high level of viability.

Taking account of these results, a complementary test was carried out in order to compare the ability of the different strains of propionic bacteria to produce significant quantities of propionic acid after having been put in contact with bile salts.

In this test 33 strains of propionic milk bacteria belonging to the TL collection of the LRTL (INRA of Rennes) were compared as to their ability to survive in the presence of bile and then to produce propionic acid:

- 20 strains belonging to the species *P. freudenreichii* subsp *shermanii*
- 6 strains belonging to the species *P. freudenreichii* subsp *freudenreichii*
- 7 strains belonging to the species *P. acidipropionici*

The operating protocol was as follows:

Starting cultures in stationary phase (2 to 3 days of culture in YEL medium incubated at 30°C) were diluted to 1/10th in YEL medium containing 0.6% of bovine bile (approximately 7-8 mmol/l of bile salts). This concentration of bile was chosen in order best to discriminate between the strains and constitutes the content of bile salts of the same order as those encountered in the duodenum.

The dilutions were incubated at 37°C for 90 minutes, then centrifuged. The bacteria were taken up in the YEL medium (initial volume) and put back to incubate at 37°C.

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After 24 hours of incubation the DO at 650 nm was measured in order to evaluate the resumption of growth. The supernatant was collected then frozen in order to determine the fatty acids.

For certain strains the experiment was repeated so as to confirm the results.

The values of the optical densities at 650 nm before the bile stress and 24 hours after the end of the stress are reported in Table 3 below:

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Table 3

Strains:	DO before the bile stress (initial DO/10)	DO at 24 hours of incubation after the bile stress	% of DO at 24 h/initial DO
<i>P. shermanii</i>			
TL125	0.32	2.02	63
TL 134	0.29 – 0.26	2.60 – 2.04	90 – 78
TL 144	0.39	3.62	93
TL 146	0.27	1.56	58
TL 147	0.28	1.23	44
TL 148	0.23	0.04	2
TL 160	0.36 – 0.32	4.67 – 4.03	130 – 126
TL 167	0.26	1.05	40
TL 168	0.32	0.57	18
TL 4	0.32	0.29	9
TL 14	0.23	0.73	32
TL 17	0.29	0.55	19
TL 22	0.28	1.39	50
TL 24	0.28	1.65	59
TL 162	0.20	2.08	104
TL 34	0.26 – 0.27	3.40 – 3.87	131 – 143
TL 50	0.26	2.05	79
TL 61	0.23	1.31	57
TL 63	0.27 – 0.26	3.26 – 3.55	121 – 137
TL 40	0.30	1.46	49
<i>P. freudenreichii</i>			
TL 142	0.34 – 0.31	2.95 – 2.80	87 – 90
TL 3	0.24	2.66	111
TL 19	0.30	2.66	89
TL 37	0.23	1.79	78
TL 33	0.26	2.38	92
TL 64	0.29	0.31	11
<i>P. acidipropionici</i>			
TL 2	0.38	2.30	61
TL 9	0.34	2.29	67
TL 15	0.34	3.09	91
TL 54	0.34	1.39	41
TL 47	0.20	0.22	11
TL 223	0.29 – 0.38	2.59 – 2.91	89 – 77
TL 249	0.44	3.40	77

It should be noted that for certain strains the final DO is higher than the initial DO, which may be explained by the growth resumed at 37°C and not at 30°C as initially.

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As a function of the results obtained, three groups of strains may be distinguished schematically:

- the strains which are distinguished from one another by a rapid resumption of growth, indicating a low mortality of the cells due to the bile (amongst them TL 34, TL 160, TL 63, TL 33, TL 15, TL 3, TL 162),
- the strains which have very low resistance to the bile, showing a very poor or zero resumption of growth (TL 148, TL 4, TL 64, TL 47),
- the intermediate strains characterised by a moderate mortality due to the bile (TL 146, TL 147, TL 167, TL 168, TL 14, TL 17, TL 22, TL 24, TL 61, TL 40, TL 54).

Only the strains having a ratio $[(DO \text{ at } 24 \text{ h}/\text{initial DO}) \times 100]$ greater than 60% (threshold chosen arbitrarily) were selected for the measurement of the lactate, of the acetate and of the propionate by HPLC in the frozen supernatants. The initial lactate content of the YEL medium was 11.4 g/l.

The concentrations of lactate, acetate and propionate of the supernatants recovered after 24 hours of incubation are set out in Table 4 below.

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Table 4

Strains:	Lactate consumed	Acetate produced	Propionate produced
	(in g/l)		
<i>P. shermanii</i>			
TL 125	5.6	0.6	2.4
TL 134	8.9 - 7.3	0.9 - 0.9	4.0 - 3.4
TL 144	8.8	1.1	4.6
TL 160	9.9 - 8.8	1.3 - 1.1	4.7 - 4.6
TL 162	5.3	0.6	2.3
TL 34	9.7 - 10.1	1.0 - 1.2	4.9 - 4.7
TL 50	6.6	0.6	3.7
TL 63	9.1 - 10.0	1.0 1.1	4.4 - 4.4
<i>P. freudenreichii</i>			
TL 142	8.1 - 8.2	0.9 - 0.9	4.5 - 4.0
TL 3	7.8	0.9	3.6
TL 19	6.6	0.7	3.7
TL 37	5.5	0.6	2.5
TL 33	7.2	0.8	3.4
<i>P. acidipropionici</i>			
TL 2	2.7	0.4	1.5
TL 9	5.3	0.6	2.4
TL 15	3.7	0.4	2.0
TL 223	2.8 - 3.8	0.4 - 0.4	1.8 - 1.7
TL 249	5.2	0.6	3.2

This table shows that in a predictable manner the quantity of propionate produced is correlated with the degree of utilisation of the lactate.

Generally speaking, the strains belonging to the species *P. acidipropionici* produce less propionic acid than the strains of the species *P. freudenreichii* in 24 hours.

In view of the results reported above, certain strains prove to be better candidates as regards production of propionate after the action of the bile. These are strains producing at least 2 g/l of propionate in the conditions described above:

TL 134, TL 50, TL 3, TL 19, TL 33, TL 249,
and preferably more than 4 g/l of propionate:

TL 160, TL 144, TL 34, TL 63, TL 142

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An experiment was also carried out on healthy volunteers with the aim of verifying the beneficial influence of gastroresistant capsules in order to improve the survival in the intestine of a strain of propionic cheese bacteria ingested in lyophilised form (TL162).

The experiment was carried out on 7 individuals in total, with 3 periods of treatment of 4 weeks, separated by intervals of 3 weeks.

Treatment 1 consisted of ingesting for 2 weeks 5×10^9 cfu/j of bacteria packaged in non-gastroresistant capsules.

Treatment 2 consisted of ingesting for 2 weeks 5×10^{10} cfu/j of bacteria packaged in non-gastroresistant capsules.

Treatment 3 consisted of ingesting for 2 weeks 5×10^9 cfu/j of bacteria packaged in gastroresistant capsules.

For each treatment 4 samples of faeces were taken in order to test for the propionic bacteria with the aid of a selective medium (Palpropionbac®, Standa-Industrie, mixed with 4 mg/l of metronidazole). The sampling dates were:

- S1: just before the period of ingestion,
- S2: 1 week after the start of ingestion,
- S3: 2 weeks after the start of ingestion,
- S4: 1 week after the end of the period of ingestion,
- HP (for the period 3): 3 weeks after the end of the period of ingestion.

During the entire experiment the volunteers could not consume cheese containing propionic bacteria in a significant quantity (Emmental, Comté, Leerdammer, Gruyères Suisses, ...) with the exception of cheese spreads.

Table 5 indicates the results of viability of the propionic bacteria in the faeces.

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With conventional capsules, the dose of 5×10^9 cfu/j (Period 1) appears insufficient in order to find viable propionic bacteria in a significant quantity in all the volunteers. On the other hand, with the dose of 5×10^{10} cfu/j (Period 2) all the volunteers had more than 5 log cfu/g of viable propionic bacteria in the faeces from the first week of treatment. However, the maximum levels of viability observed with the doses are not different (~ 7 log).

The use of gastroresistant capsules (Period 3) improves the viability of the propionic bacteria in the faeces, notably in the volunteers in whom little were found at the time of the first treatment (vol. 1, 2 and 6). In fact, in relation to Period 2 the levels of viability obtained are on average equivalent.

- on the basis of 5×10^9 cfu/j the use of gastroresistant capsules is therefore justified for a certain number of individuals (vol. 1, 2, 6), for the others they do not or only slightly improve the viability (vol. 3, 4 and 5),
- they provide results approximately equivalent to the conventional capsules containing 5×10^{10} cfu.

The short-chain fatty acids were measured in the faeces by gas chromatography. The quantities of propionate in the faeces are indicated in Table 6. For the statistical analysis, 2 groups of values were compared 2 by 2: the values corresponding to the samples of faeces where the propionic bacteria were not detected (< 4 log), for all treatments and periods combined and the values corresponding to the samples of faeces where the propionic bacteria were numbered at more than 6 log cfu/g. In the first case the average quantity of propionate is 5.06 ± 2.56 $\mu\text{mol/g}$ ($n = 25$) and in the second case it is 7.19 ± 3.18 $\mu\text{mol/g}$ ($n = 30$). These 2 values are significantly different at $p < 0.02$ (Student test). For the other short-chain fatty acids there are no significant differences. Therefore this experiment shows that the presence of substantial quantities (> 6 log cfu/g) of propionic bacteria in the colon following the ingestion of TL 162 significantly increases the quantity of propionate in the faeces. However, since the strain TL 162 does not prove to be the best candidate for optimising the production

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of propionic acid in the colon (cf. criteria for selection *in vitro*), it is probable that the results could be improved with a strain selected according to the aforementioned criteria.

Table 5 : Count of propionic bacteria in the faeces at the time of the 3 periods of treatment
(results in log cfu/g of fresh faeces)

	Vol. 1	Vol. 2	Vol. 3	Vol. 4	Vol. 5	Vol. 6	Vol. 7	average	s	n*
Period 1	S1	4,00	< 4	< 4	< 4	4,85	4,30	4,29	0,40	4/7
	S2	4,00	< 4	4,70	7,15	4,85	5,68	5,40	1,12	6/7
	S3	5,88	4,00	6,08	6,30	4,70	6,56	5,79	1,07	7/7
	S4	< 4	< 4	6,11	6,48	< 4	5,00	5,86	0,77	3/7
Period 2	S1	6,46	< 4	6,36	< 4	< 4		6,40	0,09	3/5
	S2	5,93	5,40	6,39	7,20			6,29	0,71	5/5
	S3	6,36	6,16	6,43	5,00			6,28	0,70	5/5
	S4	5,11	< 4	< 4				5,11	0,71	2/5
Period 3	S1	3,85	< 3	6,15	< 4	< 3		5,46	1,40	3/6
	S2	6,51	5,43	4,95	6,39	5,68		6,03	0,78	6/6
	S3	6,56	5,77	6,76	6,33	5,95		6,22	0,35	6/6
	S4	5,26	< 3	< 3	6,40	5,89		5,98	0,78	3/6
	HP	4,60	< 2	2,77	6,28	< 2		5,23	1,98	4/6

n* = number of individuals in which the propionic bacteria could be counted
shaded cells : all of the values higher than 6 log cfu/g.

Table 6 : Propionate concentrate in the fresh faeces (in $\mu\text{mol/g}$)

	Vol. 1	Vol. 2	Vol. 3	Vol. 4	Vol. 5	Vol. 6	Vol. 7
Period 1	S1	7,70	4,82	3,73	2,38	3,70	13,83
	S2	10,03	3,61	5,43	5,32	3,86	18,08
	S3	9,44	3,88	2,29	4,35	7,99	12,81
	S4	6,00	2,60	3,68	8,40	5,17	10,45
Period 2	S1	9,56	7,36	13,77	8,40	6,14	
	S2	7,12	1,86	5,82	5,82	4,83	
	S3	10,60	5,11	3,99	5,42		
	S4	19,16	3,12	3,58	2,16		
Period 3	S1	11,90	5,05	18,82	3,43	2,04	
	S2	9,28	8,34	7,82	5,45	7,56	
	S3	10,91	6,62	10,63	7,19	2,25	
	S4	11,93	2,89	8,91	5,74	5,20	
	HP		3,58			6,49	

shaded cells : all of the values corresponding to samples with the propionic bacteria higher than 6 log cfu/g

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Taking the foregoing into account and in accordance with a preferred characteristic of the invention, the propionic bacteria used are chosen from amongst the strains producing propionic acid in a physiologically significant quantity and in particular from amongst the strains producing at least 2 g/l of propionic acid and/or of propionates and, preferably, more than 4 g/l of propionic acid and/or of propionates after having been cultivated at 30°C in YEL medium containing approximately 11.4 g/l of lactate for 2 to 3 days, then diluted to 1/10th in a YEL medium containing 0.6% bovine bile, incubated at 37°C for 90 minutes, centrifuged, taken up in YEL medium and put back to incubate at 37°C for 24 hours.

Another selection criterion which could be taken into account in accordance with the invention corresponds to the properties of adhesion of the strains on the colonocytes: strains having good properties of adhesion in effect have the advantage of staying longer in the colon, which allows them more time to synthesise the propionic acid; moreover, the strains which become fixed can take the place of the pathogenic agents.

It should be noted that in order to obtain the effect which is sought it is not envisaged to cause the propionic acid itself to be absorbed since because of the human metabolic chain it would not be able to reach the colon and furthermore it has been shown that in high doses it is harmful to the stomach.

Amongst the beneficial effects attributed to the short-chain fatty acids synthesised at the colon and in particular to acetic acid and above all to propionic acid, one may note their role in the assimilation of the principal minerals and notably calcium iron, zinc or even magnesium; in effect it has been possible to establish that propionic acid and, to a lesser extent, acetic acid can favour the colic absorption of these minerals and the use of the absorbed fraction by the organism.

That is a particularly interesting effect in view of the fact that the assimilation of minerals is accompanied by functional effects such as by way of example the correction of anaemia for iron or bone mineralisation for calcium.

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The experimental and clinical studies which have been carried out provide a body of corroborating arguments in support of a favourable effect of propionic acid and propionates and, to a lesser extent, acetic acid and acetates on the metabolism of these minerals; this effect is probably more substantial when the conditions of digestion are poor, which leads to a substantial quantity of unabsorbed minerals being brought through the small intestine to the colic region, and when the needs are high.

The existence of correlations between the short-chain fatty acids and the metabolism of minerals has been suggested in particular by studies using soluble fibres. Thus it has been possible to establish that polysaccharides or oligosaccharides not digested by the digestive enzymes and which are therefore fermented into short-chain fatty acids (notably propionates) by the colic flora increase the absorption of minerals such as calcium, iron or zinc and that this increase is all the more marked if the conditions are pathological (deficiencies, gastrectomy ...).

Studies of colic perfusion and on the other hand the absence of effect in colectomised subjects has made it possible to confirm the localisation of the site of action at the colon.

It has also been confirmed that these actions are accompanied by a lowering of the pH and a synthesis of short-chain fatty acids; this suggests the intervention of these acids by means of a fermentation, all the more so since it has been verified that the insoluble fibres, which are not fermentable, have no effect. Furthermore, it has been established that the caecum is hypertrophied and that the colic blood flow increases, which attests to a trophic effect.

The clinical studies carried out on this subject are few, but they enable it to be confirmed that the effects of the soluble fibres are obtained by colic fermentation and show directly the effect of the short-chain fatty acids on the absorption of minerals.

Amongst these studies mention may be made of the publication *Trinidad TP, Wolever TMS, Thompson LU, Effect of acetate and propionate on calcium absorption from the rectum and distal colon of humans, Am J Clin Nutr 1996 63/ 574-578* which reports on tests in which the

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distal colon of healthy subjects was perfused directly with acetic acid, propionic acid or their association in physiological concentration; thus it was established that the disappearance of calcium from the colic orifice is increased by the short short-chain fatty acids, but in a significantly more substantial way by propionic acid; this study also showed a dose effect based on a non-saturable system of absorption. The authors suggested that the greater lipophilia of propionic acid compared with acetic acid could favour its absorption and the liberation in the colonocyte of protons whose passage into the digestive orifice would favour the absorption of calcium.

Taking account of the foregoing, the invention also relates to the use of propionic bacteria selected as a function of their nature which is not very autolytic and their ability to resist bile salts in order to obtain a current food composition or a dietetic or medicinal composition which is absorbable by a human or an animal, prepared so that the bacteria are protected at least partially against gastric acidity, containing at least 10^6 cells/gram of these bacteria, capable of favouring the assimilation of the principal minerals, in particular calcium and/or iron and/or zinc and/or magnesium at the colon.

According to a variant of the invention, it is also proposed to apply this use to obtaining a composition having antifungal properties at the colon and, in particular, capable of reducing the development there of pathogenic mycodermis of the candida/thrush type.

In fact this use takes advantage of the excellent antifungal properties of propionic acid particularly for the treatment of candidoses due to antibiotics.

It should be noted that the composition used according to the invention can, as the case may be, contain other bacteria, in particular lactic bacteria and/or bifid bacteria capable of acting in synergy with the propionic bacteria in such a way as to increase the effects mentioned above by supplying them with lactate as fermentable substrate.

The composition used according to the invention may consist of a dry or hydrated preparation presented in the form of individual fractions of approximately 100 mg to 1 g, preferably from

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200 to 500 mg, preferably containing at least 10^8 cells; in particular it may advantageously be presented in the form of gastroresistant capsules.

According to another characteristic of the invention, this composition may also consist of a formulated preparation, the propionic bacteria being added to or associated with a fermentable substrate, notably dietary fibres, or added or incorporated into liquid, paste or solid foods.

In such a preparation the propionic bacteria can play a dual role, namely technological in a first period through the fermentation of food and function in a second period since once ingested they are capable of reaching the colon and there playing the aforementioned probiotic role, particularly as regards the optimisation of the synthesis of propionic acid and the optimisation of the assimilation of minerals.

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Claims

1. Use of propionic bacteria belonging to strains which are not very autolytic and selected for their ability to produce at least 2 g/l of propionic acid and/or of propionates and preferably, more than 4 g/l of propionic acid and/or of propionates after having been cultivated at 30°C in YEL medium containing approximately 11.4 g/l of lactate for 2 to 3 days, then diluted to 1/10th in a YEL medium with 0.5% bovine bile added, incubated at 37°C for 90 minutes, centrifuged, taken up in YEL medium and put back to incubate at 37°C for 24 hours, in order to obtain a current food composition or a dietetic or medicinal composition which is absorbable by a human or an animal, prepared so that the bacteria are protected at least partially against gastric acidity, containing at least 10⁶ cells/gram of these bacteria, capable of stimulating and increasing significantly the synthesis of propionic acid and/or of propionate and, as the case may be, of acetic acid and/or of acetate at the colon by anaerobic bacterial fermentation.
2. Use as claimed in Claim 1 in order to obtain a composition capable of favouring the assimilation of the principal minerals, in particular calcium and/or iron and/or zinc and/or magnesium at the colon.
3. Use as claimed in any Claim 1, for obtaining a composition having antifungal properties at the colon and, in particular, capable of reducing the development there of pathogenic mycodermis of the candida/thrush type.
4. Use as claimed in any one of Claims 1 to 3, characterised in that the propionic bacteria belong to strains having properties of adhesion on the colonocytes.
5. Use as claimed in any one of Claims 1 to 4, characterised in that the composition consists of a dry or hydrated preparation presented in the form of individual fractions of approximately 100 mg to 1 g, preferably from 200 to 500 mg, preferably containing at least 10⁸ cells.

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6. Use as claimed in Claim 5, characterised in that the composition is presented in the form of gastroresistant capsules.
7. Use as claimed in any one of Claims 1 to 6, characterised in that the composition consists of a formulated preparation, the propionic bacteria being added to or associated with a fermentable substrate, notably dietary fibres.
8. Use as claimed in any one of Claims 1 to 6, characterised in that the composition consists of a formulated preparation, the propionic bacteria being added to or incorporated into liquid, paste or solid foods.
9. Use as claimed in any one of Claims 1 to 7, characterised in that the composition contains lactic bacteria and/or bifid bacteria.

(12) DEMANDE INTERNATIONALE PUBLIÉE EN VERTU DU TRAITÉ DE COOPÉRATION
EN MATIÈRE DE BREVETS (PCT)(19) Organisation Mondiale de la Propriété
Intellectuelle
Bureau international

TOUTES LES RECHERCHES DE BREVETS SONT FAITES EN VERTU DU TRAITÉ DE COOPÉRATION EN MATIÈRE DE BREVETS (PCT)

(43) Date de la publication internationale
25 janvier 2001 (25.01.2001)

PCT

(10) Numéro de publication internationale
WO 01/05413 A1(51) Classification internationale des brevets:
A61K 35/74, A23L 1/308, A61P 1/00, 31/00(21) Numéro de la demande internationale:
PCT/FR00/02072

(22) Date de dépôt international: 19 juillet 2000 (19.07.2000)

(25) Langue de dépôt: français

(26) Langue de publication: français

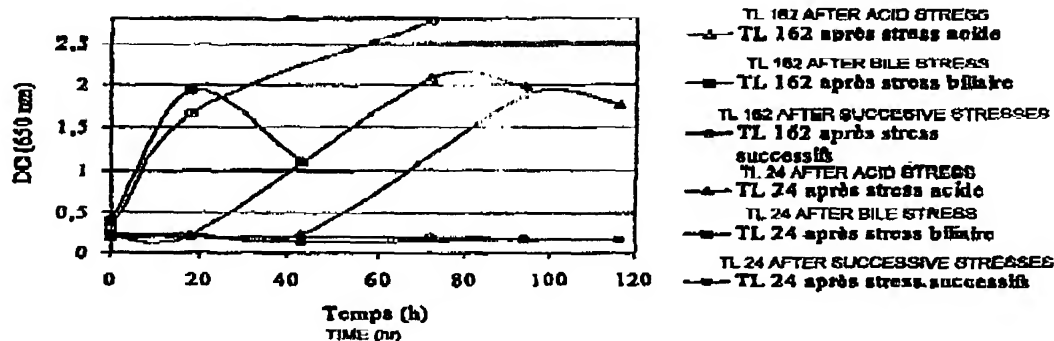
(30) Données relatives à la priorité:
99/09385 20 juillet 1999 (20.07.1999) FR(71) Déposant (pour tous les États désignés sauf US): LAB-
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vard Haussmann, F-75008 Paris (FR).(81) États désignés (national): AE, AG, AL, AM, AT, AU, AZ,
BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK,
DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID,
IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT,
LU, TV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ,

[Suite sur la page suivante]

(54) Title: USE OF PROPIONIC BACTERIA FOR PRODUCING PROPIONIC ACID AND/OR PROPIONATES IN THE COLON

(54) Titre: UTILISATION DE BACTÉRIES PROPIONIQUES POUR LA PRODUCTION D'ACIDE PROPIONIQUE ET/OU DE
PROPIONATES DANS LE COLON(57) Abstract: The invention concerns the use of propionic bacteria selected according to their low propensity to autolysis and their resistance to bile salts to obtain a current food composition or a dietetic composition of a medicine absorbable by a human or animal, prepared in such a way that the bacteria are well protected at least partially against gastric acidity, containing at least 10⁶ cell/grams of said bacteria, capable of stimulating and increasing significantly the synthesis of propionic acid and/or propionate and, as the case may be, acetic acid and/or acetate at the colon by anaerobic bacterial fermentation.(57) Abrégé: Utilisation de bactéries propioniques sélectionnées en fonction de leur caractère peu autolytique et de leur aptitude à résister aux sels biliaires pour l'obtention d'une composition d'alimentation courante ou d'une composition diététique ou médicamenteuse absorbable par l'homme ou l'animal, élaborée de façon que les bactéries soient protégées au moins partiellement vis-à-vis de l'acidité gastrique, renfermant au moins 10⁶ cellules/gramme de ces bactéries, susceptible de stimuler et d'augmenter de façon significative la synthèse d'acide propionique et/ou de propionate et le cas échéant d'acide acétique et/ou d'acétate au niveau du colon par fermentation bactérienne anaérobie.

WO 01/05413 A1

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WO 01/05413 A1

10031477 0104132

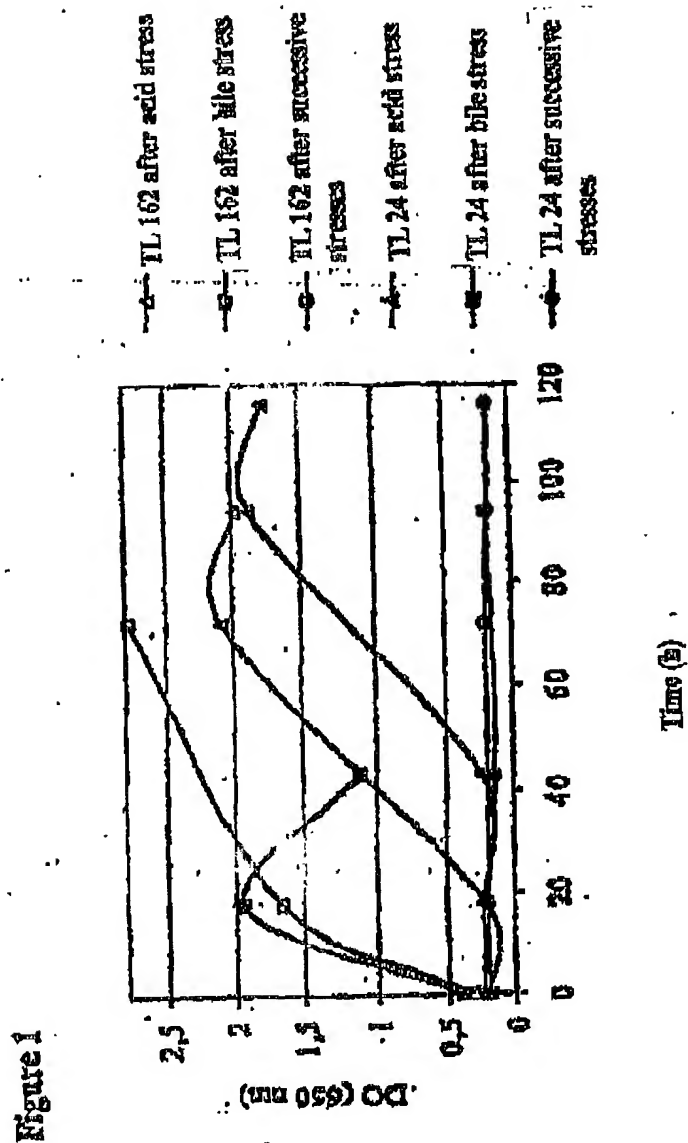
PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT,
TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.

(84) États désignés (régional): brevet ARIPO (GH, GM, KE,
LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), brevet eurasien
(AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), brevet européen
(AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU,
MC, NL, PT, SE), brevet OAPI (BF, BJ, CF, CG, CI, CM,
GA, GN, GW, ML, MR, NE, SN, TD, TG).

Publiée:

- Avec rapport de recherche internationale.
- Avant l'expiration du délai prévu pour la modification des revendications, sera republiée si des modifications sont reçues.

En ce qui concerne les codes à deux lettres et autres abrévia-
tions, se référer aux "Notes explicatives relatives aux codes et
abréviations" figurant au début de chaque numéro ordinaire de
la Gazette du PCT.



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Declaration and Power of Attorney for Patent Application

Déclaration et Pouvoirs pour Demande de Brevet

French Language Declaration

En tant que l'inventeur nommé ci-après, je déclare par le présent acte que:

Mon domicile, mon adresse postale et ma nationalité sont ceux figurant ci-dessous à côté de mon nom.

Je crois être le premier inventeur original et unique (si un seul nom est mentionné ci-dessous), ou l'un des premiers co-inventeurs originaux (si plusieurs noms sont mentionnés ci-dessous) de l'objet revendiqué, pour lequel une demande de brevet a été déposée concernant l'invention intitulée

et dont la description est fournie ci-joint à moins que la case suivante n'ait été cochée:

☐ a été déposée le _____
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_____ et modifiée le _____
_____ (le cas échéant).

Je déclare par le présent acte avoir passé en revue et compris le contenu de la description ci-dessus, revendications comprises, telles que modifiées par toute modification dont il aura été fait référence ci-dessus.

Je reconnais devoir divulguer toute information pertinente à la brevetabilité, comme défini dans le Titre 37, § 1.56 du Code fédéral des réglementations.

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

USE OF PROPIONIC BACTERIA FOR PRODUCING
PROPIONIC ACID AND/OR PROPIONATES IN THE COLON

the specification of which is attached hereto unless the following box is checked:

☒ was filed on July 19, 2000
as United States Application Number or PCT
International Application Number PCT/FR00/02072
and was amended on Jan 18, 2002; US10/031,477
(if applicable).

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, § 1.56.

French Language Declaration

Je revendique par le présent acte avoir la priorité étrangère, en vertu du Titre 35, § 119(a)-(d) ou § 365(b) du Code des Etats-Unis, sur toute demande étrangère de brevet ou certificat d'inventeur ou, en vertu du Titre 35, § 365(a) du même Code, sur toute demande internationale PCT désignant au moins un pays autre que les Etats-Unis et figurant ci-dessous et, en cochant la case, j'ai aussi indiqué ci-dessous toute demande étrangère de brevet, tout certificat d'inventeur ou toute demande internationale PCT ayant une date de dépôt précédant celle de la demande à propos de laquelle une priorité est revendiquée.

Prior foreign application(s)
Demande(s) de brevet antérieure(s)

FR 99/09385 FRANCE
(Number) (Country)
(Numéro) (Pays)

(Number) (Country)
(Numéro) (Pays)

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(Application No.)
(N° de demande)

(Filing Date)
(Date de dépôt)

(Application No.)
(N° de demande)

(Filing Date)
(Date de dépôt)

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(N° de demande)

(Filing Date)
(Date de dépôt)

(Application No.)
(N° de demande)

(Filing Date)
(Date de dépôt)

Je déclare par le présent acte que toute déclaration ci-incluse est, à ma connaissance, véridique et que toute déclaration formulée à partir de renseignements ou de suppositions est tenue pour véridique; et de plus, que toutes ces déclarations ont été formulées en sachant que toute fausse déclaration volontaire ou son équivalent est passible d'une amende ou d'une incarcération, ou des deux, en vertu de la Section 1001 du Titre 18 du Code des Etats-Unis, et que de telles déclarations volontairement fausses risquent de compromettre la validité de la demande de brevet ou du brevet délivré à partir de celle-ci.

I hereby claim foreign priority under Title 35, United States Code, § 119(a)-(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate, or § 365(a) of any PCT International application which designated at least one country other than the United States, listed below, and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or PCT International application having a filing date before that of the application on which priority is claimed.

Priority Not Claimed
Droit de priorité revendiqué

20 July 1999
(Day/Month/Year Filed)
(Jour/Mois/Année de dépôt)

(Day/Month/Year Filed)
(Jour/Mois/Année de dépôt)

I hereby claim the benefit under Title 35, United States Code, § 119(e) of any United States provisional application(s) listed below.

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States application(s), or § 365(c) of any PCT International application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, § 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application.

(Status) (patented, pending, abandoned)

(Status) (patented, pending, abandoned)
(Stato) (breveté, en cours d'examen, abandonné)

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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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POUVOIRS: En tant que l'inventeur cité, je désigne par la présente l'(les) avocat(s) et/ou agent(s) suivant(s) pour qu'ils poursuive(nt) la procédure de cette demande de brevet et traite(nt) toute affaire s'y rapportant avec l'Office des brevets et des marques: *(mentionner le nom et le numéro d'enregistrement)*.

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POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith: *(list name and registration number)*.

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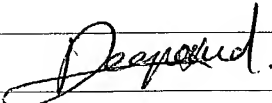
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